

University of Dundee

Keratinocyte Carcinomas

Nagarajan, Priyadharsini ; Asgari, Maryam M. ; Green, Adele C.; Guhan, Samantha M. ; Arron, Sarah T.; Proby, Charlotte

Published in:
Clinical Cancer Research

DOI:
[10.1158/1078-0432.CCR-18-1122](https://doi.org/10.1158/1078-0432.CCR-18-1122)

Publication date:
2019

Document Version
Peer reviewed version

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Nagarajan, P., Asgari, M. M., Green, A. C., Guhan, S. M., Arron, S. T., Proby, C., Rollison, D. E., Harwood, C. A., & Ewart Toland, A. (2019). Keratinocyte Carcinomas: Current concepts and future research priorities. *Clinical Cancer Research*, 25(8), 2379-2391. <https://doi.org/10.1158/1078-0432.CCR-18-1122>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Keratinocyte Carcinomas: Current concepts and future research priorities

Running Title: KCs etiology, biology and treatment

Priyadharsini Nagarajan¹, Maryam M. Asgari², Adele C. Green^{3,4}, Samantha M. Guhan², Sarah T. Arron⁵, Charlotte M. Proby⁶, Dana E. Rollison⁷, Catherine A. Harwood⁸, and Amanda Ewart Toland⁹

¹Department of Pathology, University of Texas MD Anderson Cancer Center, Houston, TX, USA

²Department of Dermatology, Massachusetts General Hospital, and Department of Population Medicine, Harvard Medical School, Boston, Massachusetts, USA

³QIMR Berghofer Medical Research Institute, Brisbane, QLD, Australia

⁴Cancer Research UK Manchester Institute and Institute of Inflammation and Repair, University of Manchester, Manchester, England, UK

⁵Department of Dermatology, University of California, San Francisco, San Francisco, California, USA

⁶Division of Cancer Research, School of Medicine, University of Dundee, Dundee, UK

⁷Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center & Research Institute, Tampa, FL.

⁸Centre for Cell Biology and Cutaneous Research, Blizard Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University, London, UK

⁹Departments of Cancer Biology and Genetics and Internal Medicine, Comprehensive Cancer Center, The Ohio State University, Columbus, OH, USA

Corresponding Author: Amanda Ewart Toland, The Ohio State University, 460 W. 12th Avenue, 998 Biomedical Research Tower, Columbus, OH, USA; Phone: 614-247-8185; Fax: 614-866-8675; E-mail: Amanda.toland@osumc.edu

Potential Conflicts of Interest: Dr. Asgari has received research grants to her institution from Valeant Pharmaceuticals and Pfizer Inc. on unrelated topics. Dr. Arron is an investigator for Leo Pharma, SunPharma, Menlo Therapeutics, Castle Biosciences, Pfizer, and Regeneron and has been a consultant for Enspectra Health, Castle Creek Pharmaceuticals, Sanofi-Genzyme, Pennside Parters, Biossance, Rakuten Aspyrian, and Genentech/Roche. Dr. Harwood has been and investigator for LeoPharma, Novartis, Galderma and PellePharm and has been a consultant for Roche and Sanofi. Dr. Proby has been an investigator for Leo Pharma and is currently an investigator for Orlucent Inc. and a partner with Emblation on an Innovate UK-funded grant.

Funding: This work was supported by the National Institutes of Health (R01 CA166672 to MA and R01 CA215151 and R21 CA219884 to AET).

Word Count: 3112, 2 Figures, 2 Tables

Abstract

Cutaneous squamous cell carcinoma (cSCC) and basal cell carcinoma (BCC) are keratinocyte carcinomas (KC), the most frequently diagnosed cancers in fair-skinned populations. Ultraviolet radiation (UVR) is the main driving carcinogen for these tumors but immunosuppression, pigmentary factors, and aging are also risk factors. Scientific discoveries have improved the understanding of the role of human papillomaviruses (HPV) in cSCC as well as the skin microbiome and a compromised immune system in the development of both cSCC and BCC. Genomic analyses have uncovered genetic risk variants, high-risk susceptibility genes, and somatic events that underlie common pathways important in KC tumorigenesis and tumor characteristics which have enabled development of prediction models for early identification of high-risk individuals. Advances in chemoprevention in high-risk individuals and progress in targeted and immune-based treatment approaches have the potential to decrease the morbidity and mortality associated with these tumors. As the incidence and prevalence of KC continue to increase, strategies for prevention, including effective sun protective behavior, educational interventions and reduction of tanning bed access and usage are essential. Gaps in our knowledge requiring additional research in order to reduce the high morbidity and costs associated with KC include better understanding of factors leading to more aggressive tumors, the roles of microbiome and HPV infection, prediction of response to therapies including immune checkpoint blockade, and how to tailor both prevention and treatment to individual risk factors and needs.

Introduction

Keratinocyte carcinoma (KC), comprised of cutaneous squamous cell carcinoma (cSCC) and basal cell carcinoma (BCC), are the most frequently diagnosed cancers in the Western world (1,2). Although the exact worldwide incidence of KC is unknown, KC represents a significant health burden in many countries. An estimated 5.4 million KC were diagnosed in the United States (US) in 2012, an increase from 3.5 million cases in 2006 (3,4). In addition to significant morbidity, they are responsible for an estimated 4000-8700 deaths per year in the US and cost ~\$4.8 billion annually (5,6). In 2014 the US Surgeon General launched the “Call to Action to Prevent Skin Cancer” which aimed to reduce skin cancer incidence and mortality, including that of both KCs and melanoma. Similar campaigns have been launched elsewhere, with most notable impact in Australia (7).

Molecular, epidemiological and clinical studies have led to greater understanding of the cellular events that occur during tumorigenesis, epidemiological risk factors, and have provided new strategies for treatment and prevention of KCs. In this review, we will discuss similarities and differences between BCC and cSCC in terms of histopathology, risk factors, and tumor development. We also highlight advances and gaps in our knowledge and emerging therapeutic and preventative strategies needed to decrease the impact of these cancers.

Overview

cSCC comprise about 20% of KC diagnoses. An estimated 3-7% of patients develop metastasis, of whom more than 70% will die from disease (8-10). BCC comprise about 80% of all KC. Despite population studies indicating that the BCC-associated mortality rate is negligible (10),

BCC can in rare cases metastasize and lead to death (11). While ratios of BCC to cSCC ranging from 2-4:1 have been reported, recent studies based on Medicare records suggest this may be changing, with equal numbers of BCCs and cSCCs being treated (3). This may reflect the aging of the population.

Risk Factors

Risk factors for KC and aggressive KC are summarized in Figure 1 and Table 1 and are detailed below. Prospective identification of high-risk patients and early intervention are facilitated by recognition of specific clinical and histopathologic characteristics for both BCC (12-17) and cSCC (9,13,18-22), so that tailored management strategies may be implemented early.

Pathophysiology

UV radiation

UV radiation is the overwhelming causative environmental carcinogen in KC. KC exhibit C>T or CC>TT dinucleotide mutations at pyrimidine bases with a strong transcription strand bias. This mutational signature (Signature 7) is characteristic of UV-induced mutation and common to almost all UV-associated skin cancers (23). KC also show a high mutational burden, far exceeding that of other cancers, although the genes mutated vary between BCCs and cSCCs (24,25). Exome sequencing of cSCC shows highest levels of *TP53* mutations and loss of function *CDKN2A* mutations. Other frequent mutations are found epigenetic regulators such as *KMT2C*, *KMT2D*, *TET2*, and loss of function Notch pathway genes such as *NOTCH1* and *NOTCH2* (24,25). Sequencing studies of metastatic cSCCs reveal higher mutational burden than primary tumors and have associated mutations in *KMT2C* with poorer outcome, including bone

metastases (25,26). Targeted sequencing revealed a high proportion of cSCCs (88%) contain potentially actionable but rare (<10%) genomic alterations including *PIK3CA*, *FGFR3*, *BRAF*, and *EGFR*, suggesting potential areas for clinical trials (27). Commonly mutated genes in BCC include those in the sonic hedgehog (SHH) signaling pathway (*PTCH1*, *SUFU*, *SMO*) as well as *TP53*. Genes mutated less frequently (8-30%) include *MYCN*, *PPP6C*, *PTPN14*, and *RBI* (28).

Immunosuppression

Innate or acquired immunosuppression is a significant risk factor for KC, particularly cSCC.

Whilst certain primary immunodeficiencies predispose to KC (29) (e.g. severe combined immunodeficiency, Wiskott-Aldrich syndrome and dyskeratosis congenita), KC are more common in acquired immunodeficiency, including immunosuppressive drug therapy (e.g. in solid-organ transplantation), immune-mediated/autoimmune inflammatory diseases (IMIDs) such as inflammatory bowel disease (IBD), vasculitis and rheumatoid arthritis (RA), non-Hodgkin lymphoma/chronic lymphocytic leukaemia (NHL/CLL) and HIV infection (30).

Solid organ transplant recipients (SOTR) are the most intensively studied iatrogenically immunosuppressed population: they have a 60-200 fold increased risk of cSCC, with reversal of the usual BCC to cSCC ratio, frequent occurrence of multiple tumors and a potentially more aggressive clinical course (31-34). Age-adjusted population estimates in the US have shown cSCC incidence ratios (IR) of 1355/100,000 person-years in SOTRs compared with 38/100,000 in the general population (35). Indeed, KC in SOTR has an IR nearly five times that of all other cancers combined in the general US population (National Cancer Institute.

<http://seer.cancer.gov/statfacts/>). Significant risk factors include age at transplantation, duration of immune suppression, skin type, gender and organ-specific factors, with greatest KC risk seen

after thoracic transplantation. In IMiDs the risk of KC is also significantly increased and this is in part treatment-related (36): exposure to thiopurines is associated with up to 5-fold increased risk for cSCC in IBD (36,37) and treatment for more than one year also increases cSCC risk in RA (38). Other non-iatrogenically immunosuppressed individuals, including those with HIV/AIDS or with hematological malignancies such as CLL, are also at significantly increased KC risk (39-41). In HIV, this risk is associated with long-term survival although highly active antiretroviral therapy may be protective (42). cSCC in association with CLL has poorer outcomes with increased recurrence and metastasis (41,43).

The pathogenesis of immunosuppression-associated KC involves a complex interplay between UVR and a number of cofactors. Innate primary and acquired immunodeficiencies are likely to result in dysregulation of tumor immune surveillance, as do immune suppressive drugs, but the latter may also contribute by direct carcinogenic effects. For example, a recent meta-analysis of 27 studies confirmed a 1.56-fold increased risk for cSCC (95% CI 1.11-2.18) in association with azathioprine (44). Thiopurines have the dual effects of causing UVA photosensitivity with consequent UVA-induced DNA damage, together with increased UVB-mutagenesis through reduced repair of UVB-induced DNA damage (45,46). A specific azathioprine signature mutation has recently been identified in cSCC (47); procarcinogenic mechanisms for the calcineurin inhibitor, cyclosporine, include reduced UV DNA damage repair (48), reduced apoptotic response to UV (49) and ATF3 induction and suppression of p53-dependent senescence (48,50). In contrast, mTOR inhibitors are associated with reduced cSCC risk, possibly through both anti-proliferative and anti-angiogenic properties (34,51-53) and the risk associated with newer immunosuppressive drugs, including tacrolimus and mycophenolate, may also be reduced, but supportive epidemiological data are not yet established (54,55).

Voriconazole, an antifungal agent commonly used in transplantation, has direct photocarcinogenic effects (56) and is associated with significantly increased risk of aggressive cSCC (57). Other drugs used in IMIDs, including anti-tumor necrosis factor agents, have also been implicated in contributing to KC risk, but data are less conclusive.

Human Papillomavirus

Patients with epidermodysplasia verruciformis (EV), a rare, autosomal recessive disorder characterized by impaired cellular immunity, represent another unique population with markedly elevated cSCC risk. Cutaneous human papillomavirus virus (cuHPV) of the genus beta (β HPV) are particularly implicated in cSCC and were first identified in patients with EV, although are also common in immunocompetent individuals (58). β HPV DNA has been detected in 18-84% of cSCCs and is three times more likely to be present in cSCCs arising among immunocompromised individuals than immunocompetent individuals (59). However, when β HPV is detected in cSCC, viral DNA is present at low copy numbers (60), and viral transcripts are absent (61). Therefore, unlike the high-risk mucosal types associated with cervical and anogenital cancers, if β HPV plays a role in keratinocyte carcinogenesis, it does so through an indirect mechanism, such as inhibition of DNA repair and/or apoptosis of UV-damaged cells (62). Multiple epidemiologic studies, incorporating both serologic and DNA-based markers of β HPV infection, have observed increased risk of cSCC associated β HPV infection (63). While these associations may simply reflect alterations in immune function that predispose individuals to both β HPV infection and cSCC, the consistent signal observed across studies underscores the need for additional research into the biology underpinning the complex interplay between UV

radiation exposure, immune function, β HPV infection and KC carcinogenesis, as β HPV vaccination could be a novel strategy for KC prevention.

Microbiome/Infection

Chronic skin diseases with altered skin microbiota such as atopic dermatitis (64), psoriasis, and hidradenitis suppurativa (65,66) may alter KC development. One study identified 6-N-hydroxyaminopurine in a strain of *S. epidermidis*, which can inhibit DNA polymerase in several human tumor cell lines, including those derived from cSCC (67). Furthermore, metagenomic analyses of the human skin microbiome revealed higher prevalence of such *S. epidermis* strains in healthy individuals. As evidence is currently circumstantial, additional studies are needed to further explore the etiopathogenic role of the microbiome in cSCC.

Germline Genetic Risk Factors and Risk Models

Although factors including immunosuppression, age, sex, pigment, and UV exposure play critical roles in the risk of developing KC (**Figure 1**), highly-penetrant pathogenic variants and lower penetrance susceptibility variants also increase risk. Hereditary syndromes associated with increased risk of cSCC are rare; these include xeroderma pigmentosa (XP), epidermolysis bullosa, Fanconi anemia, oculocutaneous albinism, and aging syndromes such as Werner syndrome (reviewed in <https://www.cancer.gov/types/skin/hp/skin-genetics-pdq>). Basal cell nevus syndrome (BCNS/Gorlin Syndrome), caused by pathogenic variants in the *PTCH1* gene and more rarely *PTCH2* (68) and *SUFU* (69), is the main syndrome associated with an increased risk of BCC. Other syndromes such as Rombo, Bazex-Dupré-Christol, and XP also show increased BCC risk (70).

Genome-wide association studies (GWAS) have identified variants (or genes) associated with increased risk for KC and melanoma. Pathways linked to increased risk of cSCC and/or BCC in the general population include genes critical for pigment (*IRF4*, *OCA2*, *HERC2*, *TYR*, *SLC45A2*, *ASIP*, *RALY*, and *MC1R*), and HLA (*HLADQA1*) (71,72). BCC GWAS have also identified variants in telomere function genes and those important in immune regulation (72). Most of these variants show small effect sizes with typical odds ratios ranging from 1.15 to 1.5. Although the total number of variants associated with KC risk is still small, there may be future benefit of using polygenic risk scores to identify individuals at elevated risk who would then be candidates for sun-protective education, behavioral intervention, and/or increased screening (73,74).

Associations between aberrant human leukocyte antigen (HLA) expression (75,76), or germline class-I and II allelic variations and KC have been controversial (77-80) and are affected by high UV exposure (81), immunosuppression (82), and HPV infection (83). Multiple variants in *HLA-DRB1* (*01, *07) have shown increased risk for BCC while *HLA-DRB1**04 was protective (82). *HLA-DRB1**01 also correlated with increased BCC risk and early tumor development in renal transplant recipients (84). Among immunosuppressed patients, class-I antigens *HLA-A03*, *HLA-A11* and *HLA-B27* and class-II antigens, *HLA-DRB1**07 and *HLA-DQA1**01 correlated with increased risk cSCC (80). GWAS analyses revealed higher cSCC risk in association with *DRB1**01, *DQA1**05:01 and *DQA1**05:05 (85), in addition to variants in *HLA-DQB1* (72), *HLA-DQA1* (71), *HLA-DRB1* (85) and *HLA-DQA1* (85). On the other hand, HLA mismatch between recipient and graft appears to have a protective effect on KC risk, with greater number of mismatched alleles conferring higher protective effect (S. Arron, manuscript under review).

Further studies may reveal the connection between HLA Class I and II antigens and KC development.

Prevention

Sun avoidance and sun protective behavior such as avoiding the sun at peak hours between 11am and 3pm, wearing protective clothing and wide-brimmed hats, regularly applying sunscreen and seeking shade have been shown in some studies to decrease the incidence of cSCC and may be effective for reduction of BCC (86,87). However, consistent adherence to these guidelines, even in high-risk populations, such as SOTRs, is suboptimal (88,89). Evidence shows that raising skin cancer awareness in high-risk populations can stimulate adoption of preventive practices (90,91) and that specific sun-protection education in specialist dermatologic-surgery clinics for SOTRs at very high KC risk, can bring about measurable behavior change (92). There remains a need for new studies to determine the delivery of effective education programs for sustained sun protective behavior strategies for prevention of KCs and to develop these to the point of regular use. Chemopreventive strategies for high-risk patients is also a consideration. The few clinical trials evaluating the effectiveness of preventive agents (e.g. tretinoin, vismodegib, nicotinamide) mostly were conducted in immunocompetent populations (93-95). Oral retinoids such as isotretinoin and acitretin, and SHH pathway inhibitor vismodegib all showed decreases in the number of BCCs in individuals with BCNS compared to placebo (94,96,97). Isotretinoin is associated with decreases in both BCCs and cSCCs in individuals with XP and in SOTRs (98,99). However, these drugs have limitations which restrict their use in the general population; for example, systemic retinoids are associated with hepatotoxicity and teratogenicity as well as xerosis, and vismodegib is associated with dysgeusia and alopecia (100). A double-blinded,

randomized controlled trial of nicotinamide (vitamin B3) in patients with a history of KCs found that 500 mg nicotinamide twice-daily reduced the incidence of BCC, cSCC and actinic keratosis compared to placebo over a 12-month period without significant side effects (93). However, there is limited evidence available for nicotinamide in OTRs in KC prevention (101), which requires confirmation in large clinical trials.

Screening

Screening the general population for KC via full body skin examination is unlikely to be cost-effective in unselected populations because specificity and accuracy of clinical diagnosis is low, and the US Preventative Service Task Force states that there is insufficient evidence to recommend KC screening for the general population (102). Increased surveillance is likely to occur resulting in increased burden on health services and costs, with unclear reduction in morbidity or mortality. On the other hand, KC screening in high-risk groups such as SOTRs may have has the potential to reduce morbidity and mortality, although there is no clear consensus on optimal screening regimens (103).

Risk models to identify individuals at highest risk for KC include sex and pigmentation, and for SOTRs, also include pre-transplant skin cancer history and age at transplant (104). Despite similarities, the different models vary in the exact factors included. The three models for SOTRs developed in small cohorts of white renal transplant recipients may not be generalizable to other populations or organ types (105-107). An ideal risk prediction tool would stratify patients based on individual factors and translate to evidence-based screening recommendations (reviewed in (104)). Implementation of existing skin cancer screening guidelines has been variable (108-112), likely reflecting availability of resources. A recent population-based study in Ontario, Canada

observed that fewer than half of SOTRs ever saw a dermatologist, but that higher adherence to annual screening after transplantation was associated with a reduction in surgically-morbid or fatal KCs (113). Economic modeling also suggests that appropriate screening and early intervention may reduce the cost of skin cancer care after transplant (114) but prospective data are needed to further justify targeted screening for reduction in KC morbidity and associated costs.

Treatment

Both BCC and cSCC can be successfully treated by a variety of modalities and guidelines for their management have been recently published (13,115-117). Treatment selection is often guided by *patient features*, such as co-morbidities and preferences, *tumor features*, that stratify KCs into low-risk and high-risk tumors (**Table 1 and Figure 1**), as well as *care features*, such as access to the modality and associated cost (118).

Surgery remains the mainstay of treatment for invasive KC and includes excision with post-operative margin assessment and Mohs micrographic surgery (MMS). Low-risk primary KCs are often treated with surgical excision whereas high-risk KCs are candidates for MMS. Non-surgical destructive options include cryosurgery, electrodesiccation and curettage (EDC), and chemical peels. EDC is widely used for low-risk KCs in non-hair bearing areas on the trunk and extremities whereas chemical peels can be used to remove superficial KCs and associated sun-damage. Light based therapies, including photodynamic therapy (PDT) and lasers, utilize discrete wavelengths of light to target KCs. Cure rates depends on tumor features, choice of photosensitizing agent, and the light source. PDT can be used to treat low-risk superficial tumors

in non-hair bearing areas. Radiation therapy is recommended for non-surgical candidates and as adjuvant treatment for tumors with extensive perineural involvement but is not recommended for patients <60 years of age or those individuals with genetic syndromes predisposing to increasing skin cancer risk. Topical treatment regimens, including 5-fluorouracil, imiquimod, ingenol mebutate, diclofenac and tazarotene, are typically reserved for superficial BCCs or SCC in-situ. Dosing regimens and cure rates vary and are impacted by the anatomic site of the tumor, side effect profiles and patient compliance. Intralesional treatment with methotrexate, 5-fluorouracil, bleomycin, or interferon is an option for patients with low-risk tumors who are not surgical candidates.

For unresectable or metastatic SCC, chemotherapeutic options have included the infusion of cisplatin, 5-FU, bleomycin, and interferon- α 2a, with low clinical response rates (<30%) (118,119). EGFR inhibition with agents including cetuximab, lapatinib, and panitumumab has shown a moderate response but their use is limited by adverse events profiles (118,120). Newer treatments, including targeted therapy for BCCs and immunotherapy and checkpoint inhibition therapy for cSCCs, hold some promise in the treatment of advanced and unresectable KCs. Currently available molecular therapies targeting the SHH signaling pathway often mutated in BCCs include vismodegib and sonidegib. Both agents have shown clinically meaningful response rates with 43% for locally advanced and 30% for metastatic disease (121-123). Their clinical utility is, limited by their side effect profile, which includes muscle spasms, alopecia, taste loss, weight loss, precluding their long-term use (121-123). Inhibition of DNA repair pathways, including PARP inhibition, is a promising future therapeutic direction for SHH pathway-resistant BCCs (124). Immune checkpoint blockade has successfully treated

hypermutated cancers, including SCC, enabling heightened sensitivity to effector T cells.

Cemiplimab, a human monoclonal antibody directed against programmed death 1, is an immune checkpoint inhibitor that has demonstrated clinical response in locally advanced (50%) and metastatic (47%) disease (125). Immune checkpoint blockade combined with other treatment modalities is a promising avenue for future systemic SCC treatment. Identification of which tumors will respond is an ongoing area of research. **Table 2** describes commonly used KC treatments and includes recommendations for use of each treatment modality (116,117,119-144).

Development of novel transdermal delivery systems such as nanoshells, sonophoresis and electroporation offer promising non-invasive alternatives for the future. Despite these advances, more data are needed to make informed decisions based on individualized risk-assessments guided by patient, tumor, and care factors. Appropriate therapeutic choice involves a shared decision-making plan that includes the provider and the patient.

Conclusions:

With increasingly longer life expectancies, the health burden associated with KCs is likely to rise still further. Our understanding of environmental risk factors such as exposure to UV radiation, immune suppression, viruses, skin microbiome, and intrinsic risk factors such as pigmentation, aging, immune function and genetic susceptibility variants on KC development is growing. However, additional research is critical in order to build on these findings, specifically to enhance sun protective behavior and public knowledge of the long-term harms of excessive UV exposure, to decrease the availability and use of indoor tanning, better capture and track KC cases via registries, improve therapies and better predict response (**Figure 2**).

Acknowledgments:

This study was supported by the National Cancer Institute of the National Institutes of Health under award numbers R01 CA166672 (to MA) and R01 CA215151 and R21 CA219884 to (AET). This content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Figure Legends:**Figure 1: Unique and shared risk factors for BCC and cSCC**

Intrinsic and extrinsic risk factors for the development of BCC and cSCC are shown including factors that are in common or unique to each tumor type.

Figure 2: Areas of Research Need

Some of the clinical and scientific areas in need of additional research to drive improvements in KC understanding, prevention, treatment, and outcomes are highlighted. IS, immunosuppression; SOTR, solid organ transplant recipients; RTR, renal transplant recipients.

Table 1. Low and high-risk features of keratinocytic carcinomas.

Features		Low-risk	High-risk	References
Basal Cell Carcinoma				
Patient	Immune status	Immunocompetent	Immunosuppressed	12, 13
Clinical	Primary vs. recurrent*	Primary	Recurrent, metastatic	13, 14, 15, 17
	Anatomic location [‡]	Area L and M	Area H	
	Site of prior radiation therapy*	No	Yes	
	Tumor dimensions*	Surface area: Area L: < 20 mm; Area M: < 10 mm Size/diameter: < 5 cm	Surface area: Area L: > 20 mm; Area M: > 10 mm Size/diameter: > 5 cm	
	Tumor circumscription*	Well-defined borders	Poorly-defined borders	
	Involvement of named nerves*	Absent	Present	
Pathologic	Histologic type / growth pattern*	Superficial, nodular, keratotic, infundibulocystic, fibroepithelioma of Pinkus	Micronodular, infiltrative, sclerosing, morpheaform, basosquamous, metatypical/sarcomatoid	13, 14, 16, 17
	Perineural invasion*	Absent	Present, diameter of involved nerve ≥ 0.1 mm, multifocality, involvement of named nerves	
Squamous Cell Carcinoma				
Patient	Immune status*	Immunocompetent	Immunosuppressed	13, 18, 19, 21
	Neurologic symptoms*	Absent	Present	
Clinical	Primary vs. recurrent*	Primary	Recurrent, metastatic	9, 13, 18, 19, 21, 22
	Anatomic location [‡]	Area L and M	Area H	
	Site of prior radiation therapy*	No	Yes	
	Site of chronic inflammation*	No	Yes	

	Rate of growth*	Slow	Rapid	
	Tumor dimensions*	Surface area: Area L: < 20 mm; Area M: < 10 mm Size/diameter: < 2 cm	Surface area: Area L: > 20 mm; Area M: > 10 mm Size/diameter: > 2 cm	
	Tumor circumscription*	Well-defined borders	Poorly-defined borders	
	Involvement of named nerves	Absent	Present	
	Extension into osseous structures	Absent	Present	
Pathologic	Histologic grade*	Well or moderately differentiated (G1-2)	Poorly differentiated (G3)	9, 13, 18-22
	Histologic type / growth pattern*	Subtype not otherwise specified	Acantholytic (adenoid), adenosquamous, desmoplastic, spindled, metaplastic/sarcomatoid	
	Perineural invasion*	Absent	Present, diameter of involved nerve \geq 0.1 mm, multifocality, involvement of deep dermal nerves or named nerves	
	Lymphovascular invasion*	Absent	Present	
	Anatomic (Clark) level*	I-III	IV-V	
	Tumor depth*	< 2.0 mm	> 2.0 mm	
	Lymph node metastasis	Absent	Present, size of metastasis > 3.0 cm, presence of extranodal extension, involvement of contralateral lymph nodes	

* Features defined by the National Comprehensive Cancer Network

ϕ Human body skin is classified into three regions according to risk for aggressive KC: area H with high-risk (frontal hair-line, central face, nose, eyelids, chin, ear, genitalia, hands, feet and bald scalp); area M with moderate-risk (cheeks, forehead, scalp, neck, jawline); and area L with low-risk (trunk and extremities, excluding H and M areas)

Table 2: Description, comparison, and efficacy, and recommended target of common KC treatments

Treatment	Description	Advantage(s)	Disadvantage(s)	Efficacy/ Recurrence Rate ³	Recommended target	References
Surgery						
Excision	Standard surgical excision followed by postoperative pathologic evaluation of margins.	<ul style="list-style-type: none"> - Lower cost than Mohs - Fast healing if surgically repaired - Allows for pathologic confirmation of tumor removal 	<ul style="list-style-type: none"> - Normal tissue not maximally conserved - May lead to substantial deformity in some anatomic sites (eyelid, nose) 	- BCC/SCC combined 5-year recurrence rate of 3.5% (CI: 1.8-5.2)	<ul style="list-style-type: none"> - Low-risk primary tumors - Select high-risk tumors with margin assessment 	116, 117, 126
Mohs	Surgical resection with intraoperative analysis of 100% of the excised margins	<ul style="list-style-type: none"> - Highest cure rate - Normal tissue maximally conserved - Allows for pathologic confirmation of tumor removal 	<ul style="list-style-type: none"> - More expensive than excision - Requires specialist to perform 	- BCC/SCC combined 5-year recurrence rate of 2.1% (0.6-3.5%)	- High-risk tumors	116, 117, 126
Destruction						
ED and C	Tumor is scraped from the skin and electricity is used to destroy remaining cancer cells in the tumor bed	<ul style="list-style-type: none"> - Minimally invasive - Cost-effective 	<ul style="list-style-type: none"> - Worse cosmetic outcome (atrophic scar) - Slow healing - Cannot be used for tumors invading fat 	<ul style="list-style-type: none"> - BCC/SCC combined 5-year recurrence rate of 4.9% (CI: 2.3-7.4%) - recurrence rates highly location and operator dependent 	- Low-risk KCs on the trunk and extremities (in non-terminal hair bearing areas)	116, 117, 126
Cryotherapy	Uses liquid nitrogen to destroys tumors cells by freeze-thaw cycles, reducing the temperature of	<ul style="list-style-type: none"> - Minimally invasive - Cost-effective - minimizes injury to normal tissue - Simple to perform 	<ul style="list-style-type: none"> - Potentially painful to patient - Worse cosmetic outcomes compared with other treatment options 	<ul style="list-style-type: none"> - BCC: 0-16.5% recurrence rate - SCC: 0.8% (CI: 0.1-2.2%) after variable follow up 	- Low-risk tumors when more effective therapies are contraindicated	116, 117, 126-128

	target tissue to -50 to -60°C					
Chemical Peels	- <i>Topical</i> solution that causes exfoliation, removing superficial KCs	- Minimally invasive	- Potential scarring (deep peels) - long recovery time - Can only be used for superficial tumors	- Long term efficacy data lacking	- Superficial primary tumors	130
Light Based Therapies						
PDT	Application of a photosensitizing agent (aminolevulinic acid-ALA or methyl aminolevulinate - MAL) which concentrate selectively in rapidly dividing cells; followed by exposure to light source, generating reactive oxygen species that destroy actively proliferating cancer cells	- Noninvasive - Selective - May be painful - Good cosmetic result	- Only recommended for superficial tumors - Treatment often not covered by insurance carriers - Requires specialized equipment - Requires training to perform - Can be costly	- BCC w/ MAL: 5-year recurrence rate of 30.7% (CI: 21.5-42.6%) - SCCs with variable follow-up: recurrence rate of 26.4% (CI: 12.3 to 43.7%)	- Primary superficial low-risk tumors	116,117, 131, 132
Lasers	- Ablative: use of a coherent light to ablate skin cancer (CO ₂ laser) - Non-ablative: selectively converts light to heat inside blood vessels (pulse dye lasers), destroying tumor	- Good cosmetic outcome (non-scarring) - Ablative lasers can also treat chronic photodamaged skin (photorejuvenation)		- BCC recurrence rate after neodymium laser treatment: 3.7% after 3 mo-5 year follow-up - SCC after neodymium laser treatment: recurrence rate of 4.4% after 3	- Resurfacing may be of benefit for those with multiple superficial, primary tumors and severe actinic damage	133, 134

				mo-5 year follow-up		
Radiation						
Traditional	<ul style="list-style-type: none"> - SXRT¹: uses high energy rays such as x-rays to destroy the KC - EBRT²: uses particles (photons, electrons or protons, most commonly electron beams) to the KC 	<ul style="list-style-type: none"> - Suitable alternative when surgery is contraindicated - Minimally invasive 	<ul style="list-style-type: none"> - Expensive - Must be performed with special equipment - Requires multiple office visits - Higher recurrence rate than surgery - causes DNA damage, increasing future KC risk 	<ul style="list-style-type: none"> - BCC 5-year recurrence rates after SXRT: 4.2% (CI: 1.9-6.4%) - SCC 5-year recurrence rates after SXRT: 5.8% (CI: 2.9-8.7%) - SCC recurrence rate after EBRT: 6.4% (CI: 3.0%-11.0%) 	<ul style="list-style-type: none"> - Low-risk tumors when surgery is not feasible or preferred - Contraindicated in genetic conditions predisposing to skin cancer (e.g., basal cell nevus syndrome, xeroderma pigmentosum) - Contraindicated in skin cancer patients with connective tissue diseases (e.g., lupus, scleroderma) - Not recommended for patients age <60 years - Need long-term data on brachytherapy 	116,117, 132, 135
Brachytherapy	Focuses X-ray radiation to the tumor with the aid of a shielded surface	<ul style="list-style-type: none"> - High dose of treatment to target tissue - Maximal sparing normal tissue - Shorter treatment times 	<ul style="list-style-type: none"> - Must be performed using special equipment - Long-term side effects include pigmentation changes, hair loss, and atrophy 	- recurrence rate varies between 0-16.7% over a period of 9 mo-10 years		136
Topical Treatment						
5-Fluorouracil	Pyrimidine analog that disrupts DNA synthesis	<ul style="list-style-type: none"> - Minimally invasive - Multiple dosing regimens 	<ul style="list-style-type: none"> - Side effects include significant local skin reactions with erythema, erosions, and crust that can last longer than a month 	Clearance rates varied by regimen and most studies lacked long term follow up.	<ul style="list-style-type: none"> - Superficial primary BCCs, not currently recommended for cSCCs based on data available 	137

			<ul style="list-style-type: none"> - Limited data regarding comparative efficacy - Imiquimod used over a large surface area can cause systemic symptoms such as the flu, fatigue, headaches, and myalgia 	<p>Clearance rates from systematic review:</p> <ul style="list-style-type: none"> - Superficial BCC: 90% - SCC in situ= 27-85% 		
Imiquimod	Stimulates the immune system through binding to toll-like receptor 7			<p>Clearance rates varied by regimen and most studies lacked long term follow up.</p> <p>Clearance rates from systematic review:</p> <ul style="list-style-type: none"> - Superficial BCC: 43-100% - Nodular BCC: 42-100% - Infiltrative BCC= 56-63% - SCC in situ= 73-88% - Invasive SCC= 71% 		137
Tazarotene	Binds to retinoid receptors, blocking the differentiation of keratinocytes			<ul style="list-style-type: none"> - BCC: complete response rate of 30.5% after 3 year follow-up - SCCIS: pilot study showed complete response of 46.6% patients after 3-5mo follow-up 		138, 139

Smoothened Inhibitors	Smoothened inhibitors (vismodegib and sonidegib) hinder HH pathway activation	- Can be used for inoperable tumors, locoregional or metastatic BCC	- Adverse events: muscle spasms, weight loss, dysgeusia, alopecia raised creatinine kinase and lipase (sonidegib), - Some BCCs develop resistance	- Median duration of response: 7.6 months - Metastatic BCC response rate: 30% (CI: 16-48%) with follow-up until 9 month after first treatment of last enrolled patient - Locally advanced BCC: 43% (CI: 31-56%) with follow-up until 9 months	Metastatic BCC or locally advanced BCC, genetic syndromes that increase BCC risk	121-123, 143
Epidermal Growth Factor Inhibitors	- EGFR is expressed by >90% of SCCs EGFR inhibitors disrupt key cellular processes - Agents used in cSCC include cetuximab, lapatinib, and panitumumab	- Can be used for inoperable tumors, locoregional or metastatic cSCC	- Side effects and systemic toxicity including acne-like rash in 78% of patients, infusions reactions, and interstitial pneumopathy	-Response rate varying from 31-69% -SCC after panitumumab: response rate of 31% with median progression free survival of 8 months -SCC after cetuximab: response rate of 69% (CI: 52-84%) after 6 weeks treatment	Metastatic cSCCs	119, 101, 144
PD-1 / PD-L1 inhibitors	- Immune checkpoint inhibition that allows T cells to	- Can be used for inoperable tumors, locoregional or metastatic cSCC	- Side effects: fatigue, nausea, constipation, rash, diarrhea; pleural	Metastatic SCC: response rate of 47% (CI: 34-61%) after	Locally advanced and metastatic cSCCs	125

	attack cancer cells (e.g. nivolumab, cemiplimab, pembrolizumab)		effusion, hypercalcemia, cellulitis, pneumonitis	median follow- up of 7.9 months	Not recommended for solid organ transplant recipients	
--	--	--	---	------------------------------------	---	--

¹ SXRT: superficial x-ray therapy

² EBRT: Electron beam radiation therapy

³ Many of these studies are small, retrospective and/or have potential selection biases so should be interpreted with caution.

References

1. Leiter U, Eigentler T, Garbe C. Epidemiology of skin cancer. *Adv Exp Med Biol* 2014;810:120-40.
2. Karimkhani C, Boyers LN, Dellavalle RP, Weinstock MA. It's time for "keratinocyte carcinoma" to replace the term "nonmelanoma skin cancer". *J Am Acad Dermatol* 2015;72:186-7.
3. Rogers HW, Weinstock MA, Feldman SR, Coldiron BM. Incidence Estimate of Nonmelanoma Skin Cancer (Keratinocyte Carcinomas) in the U.S. Population, 2012. *JAMA Dermatol* 2015;151:1081-6.
4. Rogers HW, Weinstock MA, Harris AR, Hinckley MR, Feldman SR, Fleischer AB, *et al.* Incidence estimate of nonmelanoma skin cancer in the United States, 2006. *Arch Dermatol* 2010;146:283-7.
5. Xiang F, Lucas R, Hales S, Neale R. Incidence of nonmelanoma skin cancer in relation to ambient UV radiation in white populations, 1978-2012: empirical relationships. *JAMA Dermatol* 2014;150:1063-71.
6. Guy GP, Jr., Machlin SR, Ekwueme DU, Yabroff KR. Prevalence and costs of skin cancer treatment in the U.S., 2002-2006 and 2007-2011. *Am J Prev Med* 2015;48:183-7.
7. U.S. Public Health Service OotSG. 2014 The Surgeon General's Call To Action To Prevent Skin Cancer. <<https://www.surgeongeneral.gov/library/calls/prevent-skin-cancer/call-to-action-prevent-skin-cancer.pdf>>.
8. Karia PS, Han J, Schmults CD. Cutaneous squamous cell carcinoma: estimated incidence of disease, nodal metastasis, and deaths from disease in the United States, 2012. *J Am Acad Dermatol* 2013;68:957-66.
9. Schmults CD, Karia PS, Carter JB, Han J, Qureshi AA. Factors predictive of recurrence and death from cutaneous squamous cell carcinoma: a 10-year, single-institution cohort study. *JAMA Dermatol* 2013;149:541-7.
10. Rees JR, Zens MS, Celaya MO, Riddle BL, Karagas MR, Peacock JL. Survival after squamous cell and basal cell carcinoma of the skin: A retrospective cohort analysis. *Int J Cancer* 2015;137:878-84.
11. Robinson JK, Dahiya M. Basal cell carcinoma with pulmonary and lymph node metastasis causing death. *Arch Dermatol* 2003;139:643-8.
12. Asgari MM, Moffet HH, Ray GT, Quesenberry CP. Trends in Basal Cell Carcinoma Incidence and Identification of High-Risk Subgroups, 1998-2012. *JAMA Dermatol* 2015;151:976-81.
13. Network NCC. 2017 <https://www.nccn.org/professionals/physician_gls/pdf/nmsc_blocks.pdf>.
14. Randle HW. Basal cell carcinoma. Identification and treatment of the high-risk patient. *Dermatol Surg* 1996;22:255-61.
15. Bogelund FS, Philipsen PA, Gniadecki R. Factors affecting the recurrence rate of basal cell carcinoma. *Acta Derm Venereol* 2007;87:330-4.
16. Dixon AY, Lee SH, McGregor DH. Histologic features predictive of basal cell carcinoma recurrence: results of a multivariate analysis. *J Cutan Pathol* 1993;20:137-42.
17. Welsch MJ, Troiani BM, Hale L, DelTondo J, Helm KF, Clarke LE. Basal cell carcinoma characteristics as predictors of depth of invasion. *J Am Acad Dermatol* 2012;67:47-53.
18. Nagarajan P, Ivan D. Cutaneous squamous cell carcinomas: focus on high-risk features and molecular alterations. *Glob Dermatol* 2016;3: 359-65.
19. Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: Incidence, risk factors, diagnosis, and staging. *J Am Acad Dermatol* 2018;78:237-47.
20. Liu J, Ebrahimi A, Low TH, Gao K, Palme CE, Sydney C, *et al.* Predictive value of the 8th edition American Joint Commission Cancer (AJCC) nodal staging system for patients with cutaneous squamous cell carcinoma of the head and neck. *J Surg Oncol* 2018;117:765-72.

21. Burton KA, Ashack KA, Khachemoune A. Cutaneous Squamous Cell Carcinoma: A Review of High-Risk and Metastatic Disease. *Am J Clin Dermatol* 2016;17:491-508.
22. Thompson AK, Kelley BF, Prokop LJ, Murad MH, Baum CL. Risk Factors for Cutaneous Squamous Cell Carcinoma Recurrence, Metastasis, and Disease-Specific Death: A Systematic Review and Meta-analysis. *JAMA Dermatol* 2016;152:419-28.
23. Alexandrov LB, Jones PH, Wedge DC, Sale JE, Campbell PJ, Nik-Zainal S, *et al.* Clock-like mutational processes in human somatic cells. *Nat Genet* 2015;47:1402-7.
24. South AP, Purdie KJ, Watt SA, Haldenby S, den Breems N, Dimon M, *et al.* NOTCH1 mutations occur early during cutaneous squamous cell carcinogenesis. *J Invest Dermatol* 2014;134:2630-8.
25. Yilmaz AS, Ozer HG, Gillespie JL, Allain DC, Bernhardt MN, Furlan KC, *et al.* Differential mutation frequencies in metastatic cutaneous squamous cell carcinomas versus primary tumors. *Cancer* 2017;123:1184-93.
26. Pickering CR, Zhou JH, Lee JJ, Drummond JA, Peng SA, Saade RE, *et al.* Mutational landscape of aggressive cutaneous squamous cell carcinoma. *Clin Cancer Res* 2014;20:6582-92.
27. Al-Rohil RN, Tarasen AJ, Carlson JA, Wang K, Johnson A, Yelensky R, *et al.* Evaluation of 122 advanced-stage cutaneous squamous cell carcinomas by comprehensive genomic profiling opens the door for new routes to targeted therapies. *Cancer* 2016;122:249-57.
28. Bonilla X, Parmentier L, King B, Bezrukov F, Kaya G, Zoete V, *et al.* Genomic analysis identifies new drivers and progression pathways in skin basal cell carcinoma. *Nat Genet* 2016;48:398-406.
29. Harwood CA, McGregor JM, Proby CM. Skin Cancer in the Immunocompromised Patient. In: Griffiths C, Barker J, Bleiker T, Chalmers R, Creamer D, editors. *Rook's Textbook of Dermatology*. 9 ed. Volume 4: John Wiley & Sons, Ltd. ; 2016.
30. Yanik EL, Pfeiffer RM, Freedman DM, Weinstock MA, Cahoon EK, Arron ST, *et al.* Spectrum of Immune-Related Conditions Associated with Risk of Keratinocyte Cancers among Elderly Adults in the United States. *Cancer Epidemiol Biomarkers Prev* 2017;26:998-1007.
31. Harwood CA, Mesher D, McGregor JM, Mitchell L, Leedham-Green M, Raftery M, *et al.* A surveillance model for skin cancer in organ transplant recipients: a 22-year prospective study in an ethnically diverse population. *Am J Transplant* 2013;13:119-29.
32. Lindelof B, Sigurgeirsson B, Gabel H, Stern RS. Incidence of skin cancer in 5356 patients following organ transplantation. *Br J Dermatol* 2000;143:513-9.
33. Madeleine MM, Patel NS, Plasmeijer EI, Engels EA, Bouwes Bavinck JN, Toland AE, *et al.* Epidemiology of keratinocyte carcinomas after organ transplantation. *Br J Dermatol* 2017;177:1208-16.
34. Euvrard S, Kanitakis J, Claudy A. Skin cancers after organ transplantation. *N Engl J Med* 2003;348:1681-91.
35. Garrett GL, Blanc PD, Boscardin J, Lloyd AA, Ahmed RL, Anthony T, *et al.* Incidence of and Risk Factors for Skin Cancer in Organ Transplant Recipients in the United States. *JAMA Dermatol* 2017;153:296-303.
36. Hagen JW, Pugliano-Mauro MA. Nonmelanoma Skin Cancer Risk in Patients With Inflammatory Bowel Disease Undergoing Thiopurine Therapy: A Systematic Review of the Literature. *Dermatol Surg* 2018;44:469-80.
37. Ariyaratnam J, Subramanian V. Association between thiopurine use and nonmelanoma skin cancers in patients with inflammatory bowel disease: a meta-analysis. *Am J Gastroenterol* 2014;109:163-9.
38. van den Reek JM, van Lumig PP, Janssen M, Schers HJ, Hendriks JC, van de Kerkhof PC, *et al.* Increased incidence of squamous cell carcinoma of the skin after long-term treatment with azathioprine in patients with auto-immune inflammatory rheumatic diseases. *J Eur Acad Dermatol Venereol* 2014;28:27-33.

39. Grulich AE, van Leeuwen MT, Falster MO, Vajdic CM. Incidence of cancers in people with HIV/AIDS compared with immunosuppressed transplant recipients: a meta-analysis. *Lancet* 2007;370:59-67.
40. Silverberg MJ, Leyden W, Warton EM, Quesenberry CP, Jr., Engels EA, Asgari MM. HIV infection status, immunodeficiency, and the incidence of non-melanoma skin cancer. *J Natl Cancer Inst* 2013;105:350-60.
41. Brewer JD, Habermann TM, Shanafelt TD. Lymphoma-associated skin cancer: incidence, natural history, and clinical management. *Int J Dermatol* 2014;53:267-74.
42. Zhao H, Shu G, Wang S. The risk of non-melanoma skin cancer in HIV-infected patients: new data and meta-analysis. *Int J STD AIDS* 2016;27:568-75.
43. Velez NF, Karia PS, Vartanov AR, Davids MS, Brown JR, Schmults CD. Association of advanced leukemic stage and skin cancer tumor stage with poor skin cancer outcomes in patients with chronic lymphocytic leukemia. *JAMA Dermatol* 2014;150:280-7.
44. Jiyad Z, Olsen CM, Burke MT, Isbel NM, Green AC. Azathioprine and Risk of Skin Cancer in Organ Transplant Recipients: Systematic Review and Meta-Analysis. *Am J Transplant* 2016;16:3490-503.
45. Hofbauer GF, Attard NR, Harwood CA, McGregor JM, Dziunycz P, Iotzova-Weiss G, *et al.* Reversal of UVA skin photosensitivity and DNA damage in kidney transplant recipients by replacing azathioprine. *Am J Transplant* 2012;12:218-25.
46. O'Donovan P, Perrett CM, Zhang X, Montaner B, Xu YZ, Harwood CA, *et al.* Azathioprine and UVA light generate mutagenic oxidative DNA damage. *Science* 2005;309:1871-4.
47. Inman G, Wang J, Nagano A, Alexandrov L, Purdie K, Taylor R, *et al.* The genomic landscape of cutaneous SCC reveals drivers and a novel azathioprine associated mutational signature. *Nature Communications* 2018 In press.
48. Kuschal C, Thoms KM, Boeckmann L, Laspe P, Apel A, Schon MP, *et al.* Cyclosporin A inhibits nucleotide excision repair via downregulation of the xeroderma pigmentosum group A and G proteins, which is mediated by calcineurin inhibition. *Exp Dermatol* 2011;20:795-9.
49. Norman KG, Canter JA, Shi M, Milne GL, Morrow JD, Sligh JE. Cyclosporine A suppresses keratinocyte cell death through MPTP inhibition in a model for skin cancer in organ transplant recipients. *Mitochondrion* 2010;10:94-101.
50. Wu X, Nguyen BC, Dziunycz P, Chang S, Brooks Y, Lefort K, *et al.* Opposing roles for calcineurin and ATF3 in squamous skin cancer. *Nature* 2010;465:368-72.
51. Colegio OR, Hanlon A, Olsz EB, Carucci JA. Sirolimus reduces cutaneous squamous cell carcinomas in transplantation recipients. *J Clin Oncol* 2013;31:3297-8.
52. de Fijter JW. Cancer and mTOR Inhibitors in Transplant Recipients. *Transplantation* 2017;101:45-55.
53. Hoogendijk-van den Akker JM, Harden PN, Hoitsma AJ, Proby CM, Wolterbeek R, Bouwes Bavinck JN, *et al.* Two-year randomized controlled prospective trial converting treatment of stable renal transplant recipients with cutaneous invasive squamous cell carcinomas to sirolimus. *J Clin Oncol* 2013;31:1317-23.
54. Holdaas H, De Simone P, Zuckermann A. Everolimus and Malignancy after Solid Organ Transplantation: A Clinical Update. *J Transplant* 2016;2016:4369574.
55. Rademacher S, Seehofer D, Eurich D, Schoening W, Neuhaus R, Oellinger R, *et al.* The 28-year incidence of de novo malignancies after liver transplantation: A single-center analysis of risk factors and mortality in 1616 patients. *Liver Transpl* 2017;23:1404-14.
56. Ikeya S, Sakabe JI, Yamada T, Naito T, Tokura Y. Voriconazole-induced photocarcinogenesis is promoted by aryl hydrocarbon receptor-dependent COX-2 upregulation. *Sci Rep* 2018;8:5050.

57. Mansh M, Binstock M, Williams K, Hafeez F, Kim J, Glidden D, *et al.* Voriconazole Exposure and Risk of Cutaneous Squamous Cell Carcinoma, Aspergillus Colonization, Invasive Aspergillosis and Death in Lung Transplant Recipients. *Am J Transplant* 2016;16:262-70.
58. Hampras SS, Giuliano AR, Lin HY, Fisher KJ, Abrahamsen ME, Sirak BA, *et al.* Natural history of cutaneous human papillomavirus (HPV) infection in men: the HIM study. *PLoS One* 2014;9:e104843.
59. Wang J, Aldabagh B, Yu J, Arron ST. Role of human papillomavirus in cutaneous squamous cell carcinoma: a meta-analysis. *J Am Acad Dermatol* 2014;70:621-9.
60. Weissenborn SJ, Nindl I, Purdie K, Harwood C, Proby C, Breuer J, *et al.* Human papillomavirus-DNA loads in actinic keratoses exceed those in non-melanoma skin cancers. *J Invest Dermatol* 2005;125:93-7.
61. Arron ST, Ruby JG, Dybbro E, Ganem D, Derisi JL. Transcriptome sequencing demonstrates that human papillomavirus is not active in cutaneous squamous cell carcinoma. *J Invest Dermatol* 2011;131:1745-53.
62. Tommasino M. The biology of beta human papillomaviruses. *Virus Res* 2017;231:128-38.
63. Chahoud J, Semaan A, Chen Y, Cao M, Rieber AG, Rady P, *et al.* Association Between beta-Genus Human Papillomavirus and Cutaneous Squamous Cell Carcinoma in Immunocompetent Individuals-A Meta-analysis. *JAMA Dermatol* 2016;152:1354-64.
64. Cheng J, Zens MS, Duell E, Perry AE, Chapman MS, Karagas MR. History of allergy and atopic dermatitis in relation to squamous cell and Basal cell carcinoma of the skin. *Cancer Epidemiol Biomarkers Prev* 2015;24:749-54.
65. Ring HC, Thorsen J, Saunte DM, Lilje B, Bay L, Riis PT, *et al.* The Follicular Skin Microbiome in Patients With Hidradenitis Suppurativa and Healthy Controls. *JAMA Dermatol* 2017;153:897-905.
66. Jourabchi N, Fischer AH, Cimino-Mathews A, Waters KM, Okoye GA. Squamous cell carcinoma complicating a chronic lesion of hidradenitis suppurativa: a case report and review of the literature. *Int Wound J* 2017;14:435-8.
67. Nakatsuji T, Chen TH, Butcher AM, Trzoss LL, Nam SJ, Shirakawa KT, *et al.* A commensal strain of *Staphylococcus epidermidis* protects against skin neoplasia. *Sci Adv* 2018;4:eaao4502.
68. Fan Z, Li J, Du J, Zhang H, Shen Y, Wang CY, *et al.* A missense mutation in PTCH2 underlies dominantly inherited NBCCS in a Chinese family. *J Med Genet* 2008;45:303-8.
69. Smith MJ, Beetz C, Williams SG, Bhaskar SS, O'Sullivan J, Anderson B, *et al.* Germline mutations in SUFU cause Gorlin syndrome-associated childhood medulloblastoma and redefine the risk associated with PTCH1 mutations. *J Clin Oncol* 2014;32:4155-61.
70. Castori M, Morrone A, Kanitakis J, Grammatico P. Genetic skin diseases predisposing to basal cell carcinoma. *Eur J Dermatol* 2012;22:299-309.
71. Asgari MM, Wang W, Ioannidis NM, Itnyre J, Hoffmann T, Jorgenson E, *et al.* Identification of Susceptibility Loci for Cutaneous Squamous Cell Carcinoma. *J Invest Dermatol* 2016;136:930-7.
72. Chahal HS, Wu W, Ransohoff KJ, Yang L, Hedlin H, Desai M, *et al.* Genome-wide association study identifies 14 novel risk alleles associated with basal cell carcinoma. *Nat Commun* 2016;7:12510.
73. Sordillo JE, Kraft P, Wu AC, Asgari MM. Quantifying the Polygenic Contribution to Cutaneous Squamous Cell Carcinoma Risk. *J Invest Dermatol* 2018;138:1507-10.
74. Fritsche LG, Gruber SB, Wu Z, Schmidt EM, Zawistowski M, Moser SE, *et al.* Association of Polygenic Risk Scores for Multiple Cancers in a Phenome-wide Study: Results from The Michigan Genomics Initiative. *Am J Hum Genet* 2018;102:1048-61.
75. Markey AC, Churchill LJ, MacDonald DM. Altered expression of major histocompatibility complex (MHC) antigens by epidermal tumours. *J Cutan Pathol* 1990;17:65-71.

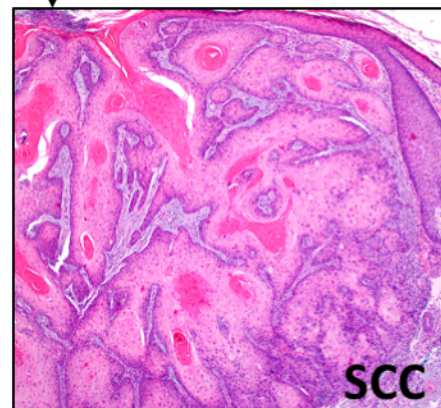
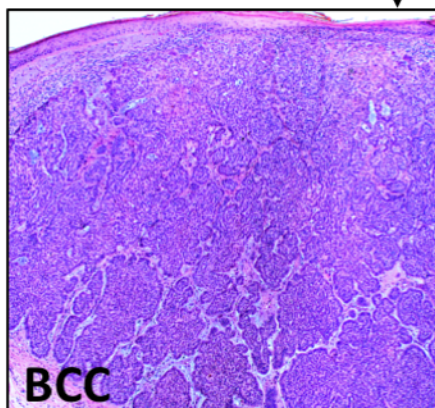
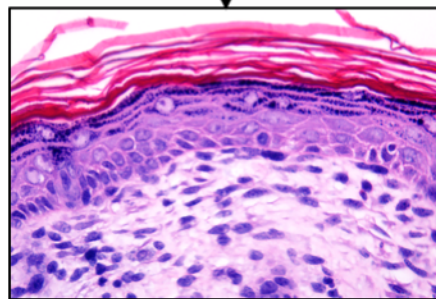
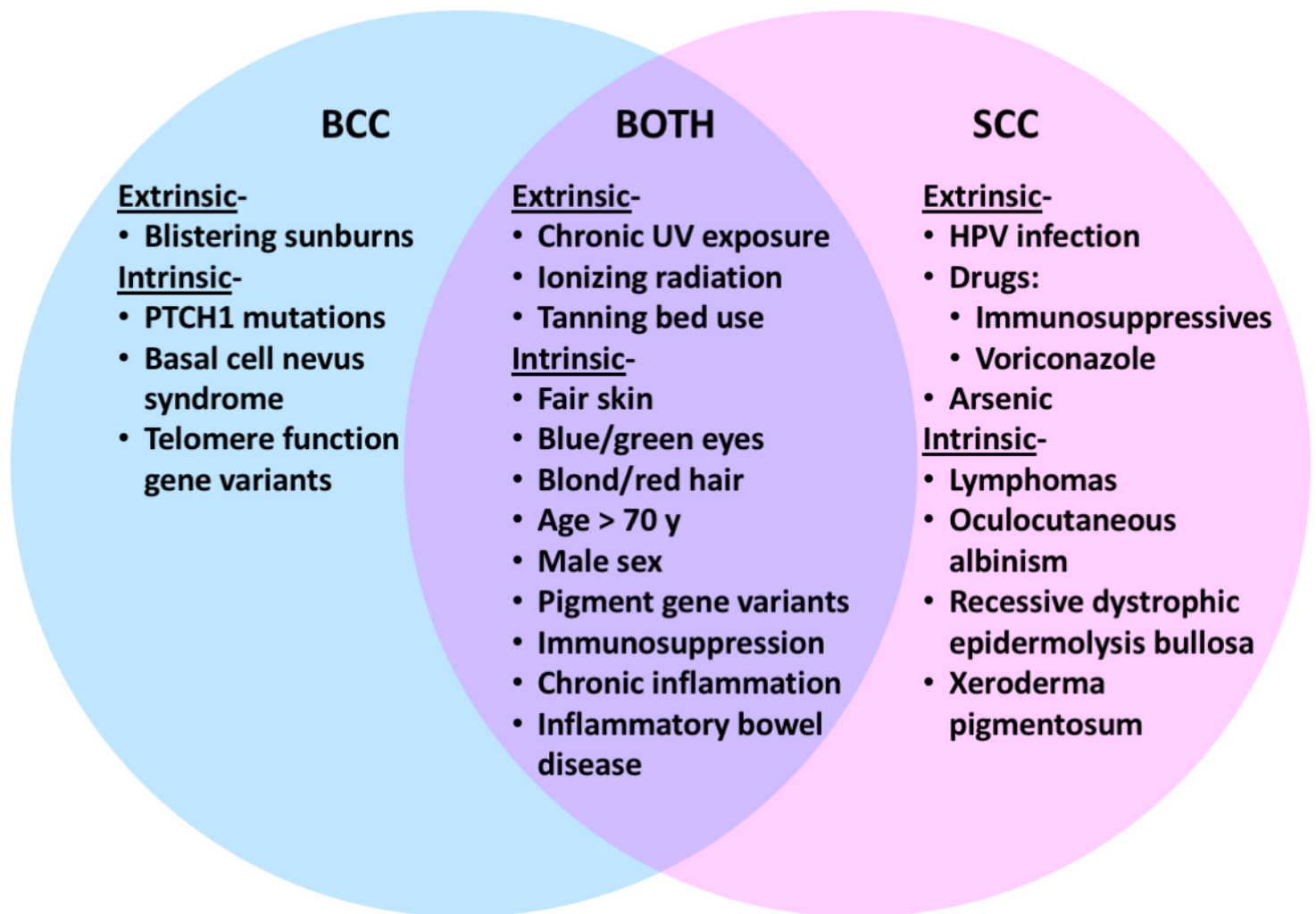
76. Mauduit G, Turbitt M, MacKie RM. Dissociation of HLA heavy chain and light chain (beta 2 microglobulin) expression on the cell surface of cutaneous malignancies. *Br J Dermatol* 1983;109:377-81.
77. Bonamigo RR, Carvalho AV, Sebastiani VR, Silva CM, Pinto AC. HLA and skin cancer. *An Bras Dermatol* 2012;87:9-16; quiz 7-8.
78. Garcia-Plata D, Mozos E, Carrasco L, Solana R. HLA molecule expression in cutaneous squamous cell carcinomas: an immunopathological study and clinical-immunohistopathological correlations. *Histol Histopathol* 1993;8:219-26.
79. Ingvar A, Ekstrom Smedby K, Lindelof B, Fernberg P, Bellocco R, Tufveson G, *et al.* No association between infections, HLA type and other transplant-related factors and risk of cutaneous squamous cell carcinoma in solid organ transplant recipients. *Acta Derm Venereol* 2012;92:609-14.
80. Yesantharao P, Wang W, Ioannidis NM, Demehri S, Whittemore AS, Asgari MM. Cutaneous squamous cell cancer (cSCC) risk and the human leukocyte antigen (HLA) system. *Hum Immunol* 2017;78:327-35.
81. Cerimele D, Contu L, Carcassi C, Costa G, La Nasa G, Sanna E, *et al.* HLA and multiple skin carcinomas. *Dermatologica* 1988;176:176-81.
82. Glover MT, Bodmer J, Bodmer W, Kennedy LJ, Brown J, Navarrete C, *et al.* HLA antigen frequencies in renal transplant recipients and non-immunosuppressed patients with non-melanoma skin cancer. *Eur J Cancer* 1993;29A:520-4.
83. Armstrong BK, Kricker A. The epidemiology of UV induced skin cancer. *J Photochem Photobiol B* 2001;63:8-18.
84. de Carvalho AV, Bonamigo RR, da Silva CM, Pinto AC. Positivity for HLA DR1 is associated with basal cell carcinoma in renal transplant patients in southern Brazil. *Int J Dermatol* 2012;51:1448-53.
85. Wang W, Ollila HM, Whittemore AS, Demehri S, Ioannidis NM, Jorgenson E, *et al.* Genetic variants in the HLA class II region associated with risk of cutaneous squamous cell carcinoma. *Cancer Immunol Immunother* 2018.
86. Green A, Whiteman D, Frost C, Battistutta D. Sun exposure, skin cancers and related skin conditions. *J Epidemiol* 1999;9:S7-13.
87. Organization WH. 2002 07/23/2018. Global solar UV index : a practical guide. WHO <<http://www.who.int/iris/handle/10665/42459>>. Accessed 2018 07/23/2018.
88. Mahe E, Morelon E, Fermanian J, Lechaton S, Pruvost C, Ducasse MF, *et al.* Renal-transplant recipients and sun protection. *Transplantation* 2004;78:741-4.
89. Iannacone MR, Pandeya N, Isbel N, Campbell S, Fawcett J, Soyer HP, *et al.* Sun Protection Behavior in Organ Transplant Recipients in Queensland, Australia. *Dermatology* 2015;231:360-6.
90. Wu SZ, Jiang P, DeCaro JE, Bordeaux JS. A qualitative systematic review of the efficacy of sun protection education in organ transplant recipients. *J Am Acad Dermatol* 2016;75:1238-44 e5.
91. Holman DM, Ding H, Guy GP, Jr., Watson M, Hartman AM, Perna FM. Prevalence of Sun Protection Use and Sunburn and Association of Demographic and Behavioral Characteristics With Sunburn Among US Adults. *JAMA Dermatol* 2018;154:561-8.
92. Papier K, Gordon LG, Khosrotehrani K, Isbel N, Campbell S, Griffin A, *et al.* Increase in preventive behaviour by organ transplant recipients after sun protection information in a skin cancer surveillance clinic. *Br J Dermatol* 2018.
93. Chen AC, Martin AJ, Choy B, Fernandez-Penas P, Dalziel RA, McKenzie CA, *et al.* A Phase 3 Randomized Trial of Nicotinamide for Skin-Cancer Chemoprevention. *N Engl J Med* 2015;373:1618-26.

94. Tang JY, Ally MS, Chanana AM, Mackay-Wiggan JM, Aszterbaum M, Lindgren JA, *et al.* Inhibition of the hedgehog pathway in patients with basal-cell nevus syndrome: final results from the multicentre, randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Oncol* 2016;17:1720-31.
95. Weinstock MA, Bingham SF, Digiovanna JJ, Rizzo AE, Marcolivio K, Hall R, *et al.* Tretinoin and the prevention of keratinocyte carcinoma (Basal and squamous cell carcinoma of the skin): a veterans affairs randomized chemoprevention trial. *J Invest Dermatol* 2012;132:1583-90.
96. Tang JY, Mackay-Wiggan JM, Aszterbaum M, Yauch RL, Lindgren J, Chang K, *et al.* Inhibiting the hedgehog pathway in patients with the basal-cell nevus syndrome. *N Engl J Med* 2012;366:2180-8.
97. Chang AL, Solomon JA, Hainsworth JD, Goldberg L, McKenna E, Day BM, *et al.* Expanded access study of patients with advanced basal cell carcinoma treated with the Hedgehog pathway inhibitor, vismodegib. *J Am Acad Dermatol* 2014;70:60-9.
98. Bavinck JN, Tieben LM, Van der Woude FJ, Tegzess AM, Hermans J, ter Schegget J, *et al.* Prevention of skin cancer and reduction of keratotic skin lesions during acitretin therapy in renal transplant recipients: a double-blind, placebo-controlled study. *J Clin Oncol* 1995;13:1933-8.
99. Kraemer KH, DiGiovanna JJ, Moshell AN, Tarone RE, Peck GL. Prevention of skin cancer in xeroderma pigmentosum with the use of oral isotretinoin. *N Engl J Med* 1988;318:1633-7.
100. Xie P, Lefrancois P. Efficacy, Safety, and Comparison of Sonic Hedgehog Inhibitors in Basal Cell Carcinomas: A Systematic Review and Meta-Analysis. *J Am Acad Dermatol* 2018.
101. Chen AC, Martin AJ, Dalziel RA, McKenzie CA, Lowe PM, Eris JM, *et al.* A phase II randomized controlled trial of nicotinamide for skin cancer chemoprevention in renal transplant recipients. *Br J Dermatol* 2016;175:1073-5.
102. Force USPST, Bibbins-Domingo K, Grossman DC, Curry SJ, Davidson KW, Ebell M, *et al.* Screening for Skin Cancer: US Preventive Services Task Force Recommendation Statement. *JAMA* 2016;316:429-35.
103. Acuna SA, Huang JW, Scott AL, Micic S, Daly C, Brezden-Masley C, *et al.* Cancer Screening Recommendations for Solid Organ Transplant Recipients: A Systematic Review of Clinical Practice Guidelines. *Am J Transplant* 2017;17:103-14.
104. Lowenstein SE, Garrett G, Toland AE, Jambusaria-Pahlajani A, Asgari MM, Green A, *et al.* Risk prediction tools for keratinocyte carcinoma after solid organ transplantation: a review of the literature. *Br J Dermatol* 2017;177:1202-7.
105. Carroll RP, Ramsay HM, Fryer AA, Hawley CM, Nicol DL, Harden PN. Incidence and prediction of nonmelanoma skin cancer post-renal transplantation: a prospective study in Queensland, Australia. *Am J Kidney Dis* 2003;41:676-83.
106. Harden P, Fryer A, Reece S, Smith A, Ramsay H. Annual incidence and predicted risk of nonmelanoma skin cancer in renal transplant recipients. *Transplantation Proceedings* 2001;33:1302-4.
107. Urwin H, Jones P, Harden P, Ramsay H, Hawley C, Nicol D, *et al.* Predicting risk of nonmelanoma skin cancer and premalignant skin lesions in renal transplant recipients. *Transplantation* 2009;87:1667-71.
108. Cowen EW, Billingsley EM. Awareness of skin cancer by kidney transplant patients. *J Am Acad Dermatol* 1999;40:697-701.
109. Horn J, Lock-Andersen J, Rasmussen K, Jemec GB. [Screening for skin cancer in organ transplant recipients in Denmark]. *Ugeskr Laeger* 2005;167:2762-5.
110. Thurot-Guillou C, Templier I, Janbon B, Pinel N, Beani JC, Leccia MT. [Dermatologic follow-up and evaluation of skin tumours in renal transplant patients]. *Ann Dermatol Venerol* 2007;134:39-44.

111. Garg S, Carroll RP, Walker RG, Ramsay HM, Harden PN. Skin cancer surveillance in renal transplant recipients: re-evaluation of U.K. practice and comparison with Australian experience. *Br J Dermatol* 2009;160:177-9.
112. Lloyd A, Klintmalm G, Qin H, Menter A. Skin cancer evaluation in transplant patients: a physician opinion survey with recommendations. *Clin Transplant* 2015;29:110-7.
113. Chan AW, Fung K, Austin PC, Kim SJ, Singer LG, Baxter NN, *et al.* Improved keratinocyte carcinoma outcomes with annual dermatology assessment after solid organ transplantation: Population-based cohort study. *Am J Transplant* 2018.
114. Ruegg CP, Graf N, Muhleisen B, Szucs TD, French LE, Surber C, *et al.* Squamous cell carcinoma of the skin induces considerable sustained cost of care in organ transplant recipients. *J Am Acad Dermatol* 2012;67:1242-9.
115. Kim JYS, Kozlow JH, Mittal B, Moyer J, Olencki T, Rodgers P. Guidelines of care for the management of cutaneous squamous cell carcinoma. *J Am Acad Dermatol* 2018;78:560-78.
116. Work G, Invited R, Kim JYS, Kozlow JH, Mittal B, Moyer J, *et al.* Guidelines of care for the management of basal cell carcinoma. *J Am Acad Dermatol* 2018;78:540-59.
117. Work G, Invited R, Kim JYS, Kozlow JH, Mittal B, Moyer J, *et al.* Guidelines of care for the management of cutaneous squamous cell carcinoma. *J Am Acad Dermatol* 2018;78:560-78.
118. Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: Management of advanced and high-stage tumors. *J Am Acad Dermatol* 2018;78:249-61.
119. Cranmer LD, Engelhardt C, Morgan SS. Treatment of unresectable and metastatic cutaneous squamous cell carcinoma. *Oncologist* 2010;15:1320-8.
120. Maubec E, Petrow P, Scheer-Senyarich I, Duvillard P, Lacroix L, Gelly J, *et al.* Phase II study of cetuximab as first-line single-drug therapy in patients with unresectable squamous cell carcinoma of the skin. *J Clin Oncol* 2011;29:3419-26.
121. Sekulic A, Migden MR, Oro AE, Dirix L, Lewis KD, Hainsworth JD, *et al.* Efficacy and safety of vismodegib in advanced basal-cell carcinoma. *N Engl J Med* 2012;366:2171-9.
122. Basset-Seguin N, Hauschild A, Grob JJ, Kunstfeld R, Dreno B, Mortier L, *et al.* Vismodegib in patients with advanced basal cell carcinoma (STEVE): a pre-planned interim analysis of an international, open-label trial. *Lancet Oncol* 2015;16:729-36.
123. Migden MR, Guminski A, Gutzmer R, Dirix L, Lewis KD, Combemale P, *et al.* Treatment with two different doses of sonidegib in patients with locally advanced or metastatic basal cell carcinoma (BOLT): a multicentre, randomised, double-blind phase 2 trial. *Lancet Oncol* 2015;16:716-28.
124. Chiang A, Jaju PD, Batra P, Rezaee M, Epstein EH, Jr., Tang JY, *et al.* Genomic Stability in Syndromic Basal Cell Carcinoma. *J Invest Dermatol* 2018;138:1044-51.
125. Migden MR, Rischin D, Schmults CD, Guminski A, Hauschild A, Lewis KD, *et al.* PD-1 Blockade with Cemiplimab in Advanced Cutaneous Squamous-Cell Carcinoma. *N Engl J Med* 2018; 379:341-351.
126. Chren MM, Linos E, Torres JS, Stuart SE, Parvataneni R, Boscardin WJ. Tumor recurrence 5 years after treatment of cutaneous basal cell carcinoma and squamous cell carcinoma. *J Invest Dermatol* 2013;133:1188-96.
127. Holt PJ. Cryotherapy for skin cancer: results over a 5-year period using liquid nitrogen spray cryosurgery. *Br J Dermatol* 1988;119:231-40.
128. Kuflik EG, Gage AA. The five-year cure rate achieved by cryosurgery for skin cancer. *J Am Acad Dermatol* 1991;24:1002-4.
129. Thissen MR, Neumann MH, Schouten LJ. A systematic review of treatment modalities for primary basal cell carcinomas. *Arch Dermatol* 1999;135:1177-83.
130. Kaminaka C, Yamamoto Y, Yonei N, Kishioka A, Kondo T, Furukawa F. Phenol peels as a novel therapeutic approach for actinic keratosis and Bowen disease: prospective pilot trial with

- assessment of clinical, histologic, and immunohistochemical correlations. *J Am Acad Dermatol* 2009;60:615-25.
131. Roozeboom MH, Aardoom MA, Nelemans PJ, Thissen MR, Kelleners-Smeets NW, Kuijpers DI, *et al.* Fractionated 5-aminolevulinic acid photodynamic therapy after partial debulking versus surgical excision for nodular basal cell carcinoma: a randomized controlled trial with at least 5-year follow-up. *J Am Acad Dermatol* 2013;69:280-7.
 132. Lansbury L, Bath-Hextall F, Perkins W, Stanton W, Leonardi-Bee J. Interventions for non-metastatic squamous cell carcinoma of the skin: systematic review and pooled analysis of observational studies. *BMJ* 2013;347:f6153.
 133. Choudhary S, Tang J, Elsaie ML, Nouri K. Lasers in the treatment of nonmelanoma skin cancer. *Dermatol Surg* 2011;37:409-25.
 134. Moskalik K, Kozlov A, Demin E, Boiko E. The efficacy of facial skin cancer treatment with high-energy pulsed neodymium and Nd:YAG lasers. *Photomed Laser Surg* 2009;27:345-9.
 135. Cognetta AB, Howard BM, Heaton HP, Stoddard ER, Hong HG, Green WH. Superficial x-ray in the treatment of basal and squamous cell carcinomas: a viable option in select patients. *J Am Acad Dermatol* 2012;67:1235-41.
 136. Delishaj D, Rembielak A, Manfredi B, Ursino S, Pasqualetti F, Laliscia C, *et al.* Non-melanoma skin cancer treated with high-dose-rate brachytherapy: a review of literature. *J Contemp Brachytherapy* 2016;8:533-40.
 137. Love WE, Bernhard JD, Bordeaux JS. Topical imiquimod or fluorouracil therapy for basal and squamous cell carcinoma: a systematic review. *Arch Dermatol* 2009;145:1431-8.
 138. Bianchi L, Orlandi A, Campione E, Angeloni C, Costanzo A, Spagnoli LG, *et al.* Topical treatment of basal cell carcinoma with tazarotene: a clinicopathological study on a large series of cases. *Br J Dermatol* 2004;151:148-56.
 139. Bardazzi F, Bianchi F, Parente G, Guareschi E, Landi C. A pilot study on the use of topical tazarotene to treat squamous cell carcinoma in situ. *J Am Acad Dermatol* 2005;52:1102-4.
 140. Siller G, Rosen R, Freeman M, Welburn P, Katsamas J, Ogbourne SM. PEP005 (ingenol mebutate) gel for the topical treatment of superficial basal cell carcinoma: results of a randomized phase IIa trial. *Australas J Dermatol* 2010;51:99-105.
 141. Brinkhuizen T, Frencken KJ, Nelemans PJ, Hoff ML, Kelleners-Smeets NW, Zur Hausen A, *et al.* The effect of topical diclofenac 3% and calcitriol 3 mug/g on superficial basal cell carcinoma (sBCC) and nodular basal cell carcinoma (nBCC): A phase II, randomized controlled trial. *J Am Acad Dermatol* 2016;75:126-34.
 142. Kirby JS, Miller CJ. Intralesional chemotherapy for nonmelanoma skin cancer: a practical review. *J Am Acad Dermatol* 2010;63:689-702.
 143. Pricl S, Cortelazzi B, Dal Col V, Marson D, Laurini E, Fermeglia M, *et al.* Smoothed (SMO) receptor mutations dictate resistance to vismodegib in basal cell carcinoma. *Mol Oncol* 2015;9:389-97.
 144. Foote MC, McGrath M, Guminski A, Hughes BG, Meakin J, Thomson D, *et al.* Phase II study of single-agent panitumumab in patients with incurable cutaneous squamous cell carcinoma. *Ann Oncol* 2014;25:2047-52.

RISK FACTORS FOR KERATINOCYTIC CANCERS



KERATINOCYTE CARCINOMAS

ETIOLOGY

Regulation of immune surveillance
Pathways leading to metastasis
Role of viruses and microbiome
Epidemiology and KC registries

RISK PREDICTION

Models to predict KC
Polygenic risk scores
Role of IS minimization in SOTRs

THERAPIES

Individualized treatment strategies
Targeted mutation-based therapies
Predictors of response for immunomodulators and checkpoint inhibitors
Role of IS inhibitors in RTRs

SCREENING

Better predictors of aggressive tumors
Prospective screening for KC

PRIMARY PREVENTION IN HIGH RISK POPULATIONS

Nicotinamide in SOTRs
HPV vaccines
Promoting sun protective behavior